

# GRAPE INSECTS

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The world's vineyards cover 10 million hectares and produce 250 million hectolitres of wine, 70 million hundredweight of table grapes, 9 million hundredweight of dried grapes, and 2.5 million hundredweight of concentrate. Thus, both in terms of quantities produced and the value of its products, the vine constitutes a particularly important cultivation.

## THE HOST PLANT AND ITS CULTIVATION

The original area of distribution of the genus *Vitis* was broken up by the separation of the continents; although numerous species developed, *Vitis vinifera* has been cultivated from the beginning for its fruit and wine producing qualities (43, 75, 184). This cultivation commenced in Transcaucasia about 6000 B.C. Subsequent human migration spread its cultivation, at first around the Mediterranean coast; the Roman conquest led to the plant's progressive establishment in Europe, almost to its present extent. Much later, the Western Europeans planted the grape vine wherever cultivation was possible, i.e. throughout the temperate and warm temperate regions of the world: North America, particularly California; South America, North Africa, South Africa, Australia, etc.

Since the commencement of vine cultivation, man has attempted to increase its production, both in terms of quality and quantity, by various means including selection of mutations or hybridization. A wide variety of cultivars have been obtained and then spread by vegetative multiplication (74).

Viticulturists, in wishing to spread the cultivation of the vine, have had to choose among the numerous varieties (clones) those that were best adapted to the soil or climate of each particular region. Moreover, a variety of ecological conditions (e.g. soil and climate) have led the viticulturists to carry out many different cultivation practices. Principally because of pruning the vine has become an extraordinary polymorphic plant. Low pruning results in rootstocks bearing short shoots that are not more than 50-60 cm high (bush, cordon, or fan pruning), whereas the tall vines

(trellised, arbor, trained vines, and vines on trees) can reach a height of several meters. Moreover, modern methods of cultivation, particularly mechanized harvesting, have caused the creation of *wide and tall vines*—rows of widely separated rootstocks (2.5 m or more apart), as planned vegetation.

This polymorphism and the diversity of plantation methods creates ecological conditions which inhibit or favor the action of the animal or vegetable pests of the vine. It is highly probable that the modern procedures of noncultivation (suppression of ploughing and use of herbicides) will soon have an influence on the action of pests.

## THE PESTS<sup>1</sup>

Originating from a rather limited region, grapes are now intensively cultivated, sometimes to the extent of being a monoculture. When growing over many tens of thousands of hectares, the vine has attracted diverse pests indigenous to the ecosystems into which it has been introduced (4). These pests have adapted so well to the plant that occasionally biological races infesting the vine have developed from indigeneous species. Less commonly, monophagous insects from other *Vitis* species have been transported on plant samples and have infested *V. vinifera*. This is the case with Phylloxera, an insect whose appearance in Europe considerably disturbed and subsequently modified the cultivation of the vine there.

An important note must also be added on the subject of the adaptation of indigeneous insects during the introduction of the vine into a given country. The ecological niches offered by the biocenosis of the vine has been occupied in each country by different species. For example, the vine chrysomelids (*écrivains*) in Europe are different species from those in North America. It is the same for the so-called grape worms. However, these species quite often belong to the same genus—*Lobesia botrana* in Europe, *Lobesia viteana* in the USA. Consequently, pests of the vine will be considered in relation to the part of the plant attacked (Table 1).

On account of the large number of pests attacking the plant throughout the world, papers that provide an exhaustive list are rare; in fact, the only one available is that by Stellwaag in 1928 (174). However, monographs on vine pests are easily found in each vine-growing country: Spain (48), Italy (23, 168), Algeria (62), Switzerland (41), Romania (121), Moldavia (166, 201), Turkey (80, 88), Azerbaidjan (100), USSR (114), California (170), eastern USA (66, 123), Chile (137), Japan (89), Australia (18, 125, 182), and France (19, 20, 32, 43, 64, 75, 110, 122).

### *Root Pests*

Formerly the most damaging pest of the vine was, without doubt, Phylloxera, *Dactylospheera (Viteus) vitifolii*, which in a few years completely destroyed 2,500,000 ha of vine in western Europe. The species was described in 1854 in New York State and subsequently recorded at several sites in the eastern United States.

<sup>1</sup>Only insects and mites are studied here; nematodes are not, even though they cause direct damage to the vine roots and even though certain species of the genus *Xiphinema*, for example, are redoubtable vectors of virus (82, 198).

Table 1 Grape insects

Plant organ attacked	Pest		Ref.
	Order	Region	
Roots	Homoptera		
	<i>Dactylospheera (Viteus) vitifolii</i>	World	12, 33-36, 42, 58, 98, 99, 107, 108, 117, 118, 122, 134, 135, 170, 175-179, 186, 187
	<i>Cicada</i> spp.	S. Europe	174
	<i>Tibicen haematodes</i>	S. Europe	29
	<i>Cicada septemdecim</i>	California	170
	<i>Rhizoecus falcifer</i>	California	170
	<i>Margarodes meridionalis</i>	California	170
	<i>Margarodes vitis</i>	Chili	174
	<i>Margarodes capensis</i>	S. Africa	174
	<i>Margarodes greeni</i>	S. Africa	174
	<i>Eurhizoecus brasiliensis</i>	Brasil	72
	Coleoptera		
	<i>Bromius obscurus</i>	Europe, N. America	19
	<i>Fidia viticida</i>	N. America	66
	<i>Scelodonta strigicollis</i>	India	109
	<i>Vesperus</i> spp.	France, Spain, Italy	63, 146
	<i>Pentodon</i> spp.	S. Europe	19, 32, 122
	<i>Phyllognatus excavatus</i>	S. Europe	85
	<i>Opatrum sabulosum</i>	Europe	32
	Lepidoptera		
<i>Vitacea polistiformis</i>	Missouri	59, 155	
Wood	Trunk	T. Isoptera	
		<i>Caloterme flavicollis</i>	S. Europe 71
		<i>Reticulitermes lucifugus</i>	S. Europe 71
	<i>Reticulitermes hesperus</i>	California 170	
	Lepidoptera		
	<i>Cossus cossus</i>	S. Europe 70	
	<i>Paropta paradoxus</i>	Israel, Egypt 141	
	Coleoptera		
	<i>Anaglyptus mysticus</i>	Bulgaria 163	
	Shoots	S. Homoptera	
		<i>Eulecanium corni</i>	S. Europe 20, 32
		<i>Pulvinaria vitis</i>	S. Europe 20, 32
		<i>Diaspidiotus uvae</i>	California 170
		Divers	
		<i>Ceresa bubalus</i> (Membracidae)	E. United States 143, 168, 192
Bostrychidae		S. Europe 122, 129	
<i>Macrophya strigosa</i> (Tenthredinidae)		S. Europe 122	
<i>Polycaon confertus</i> (Coleoptera)	170		
Buds and very young shoots	Lepidoptera		
	<i>Arctia caja</i>	S. Europe 19, 20, 27, 32	
	Noctuidae	World 19, 20, 26, 32, 53, 151, 170	
	Coleoptera		
	<i>Peritelus sphaeroides</i>	S. Europe 19, 32, 84	
	<i>Peritelus noxius</i>	S. Europe 19, 32, 84	

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Table 1 (Continued)

Plant organ attacked	Pest		Ref.
	Order	Region	
	<i>Limonus canus</i>	California	170
	<i>Glyptoscelis squamulata</i>	California	170
	<i>Pocalta ursina</i>	California	170
	<i>Phlyctinus callosus</i>	S. Africa	24, 25
	<i>Eremnus cerealis</i>	S. Africa	24, 25
	<i>Eremnus stulosus</i>	S. Africa	24, 25
	Acarina		
	<i>Calepitrimerus vitis</i>	S. Europe	120, 148
	<i>Eriophyes oculivitis</i>	Egypt	11
	<i>Eriophyes vitineusgemma</i>	Moldavia	119
Leaf	Lepidoptera		
	<i>Sparganothis pilleriana</i>	S. Europe	142, 145, 149, 169
	<i>Celerio lineata</i>	Europe, N. America	19, 20, 32, 170
	<i>Pholus achemon</i>	California	170
	<i>Antispila rivillei</i>	Georgia	57
	<i>Harrisina brillans</i>	Mexico, California	84a, 123, 140, 170
	<i>Sylepta lunalis</i>	India	136
	Coleoptera		
	<i>Haltica lythri</i> subsp. <i>ampelophaga</i>	Europe	122
	<i>Haltica chalybea</i>	California	170
	<i>Haltica torquata</i>	California	170
	<i>Byctiscus betulae</i>	Europe	19
	<i>Desmia funeralis</i>	N. America	5, 69, 93
	Homoptera		
	<i>Aphis illinoisensis</i>	E. United States	123
	Cicadoidea		
	<i>Philaenus spumarius</i>	N. America, Europe	83, 144, 170, 200
	<i>Scaphoideus littoralis</i>	N. America, France, Germany, Switzerland, Italy	45, 50, 51, 76, 79
	<i>Empoasca flavescens</i>	Europe	13, 14, 30, 31, 52, 57, 128, 131, 160, 162, 189, 190, 193
	<i>Empoasca lybica</i>	Spain, S. Italy, Maghreb, Tanganyka	191
	<i>Flata ferrugata</i>	Punjab	171
	<i>Unnata intracta</i>	Punjab	171
	<i>Zygina rhamnii</i>	France	
	<i>Erythroneura adanae vitisuga</i>	Bulgaria	102
	<i>Erythroneura comes</i>	California	96, 170, 173
	<i>Erythroneura variabilis</i>	California	96, 170, 173
	<i>Erythroneura elegantulae</i>	California	96, 170, 173
	<i>Erythroneura ziczac</i>	British Columbia	124
	Heteroptera		
	<i>Nyzius senecionis</i>	France	
	<i>Nyzius ericae</i>	Europe, America	22
	Acarina		
	<i>Eotetranychus carpini</i>	S. Europe	65, 81
	<i>Panonychus ulmi</i>	S. Europe	47
	<i>Brevipalpus lewisi</i>	Bulgaria	21

Table 1 (Continued)

Plant organ attacked	Pest		Ref.
	Order	Region	
Gall makers	<i>Tetranychus pacificus</i>	California	104, 105, 111, 173
	<i>Tetranychus flavus</i>	California	170, 173
	<i>Tetranychus atlanticus</i>	France	
	<i>Oligonychus mangiferae</i>	India	167
	Acarina		
	<i>Eriophyes vitis</i>	S. Europe	77, 126, 148
	<i>Eriophyes vitigenusgemma</i>	Moldavia	119
	<i>Eriophyes oculivitis</i>	Egypt	11, 202
	Homoptera		
	<i>Dactylospheera (Viteus) vitifolii</i>	World	179, 180
Fruits	Diptera		
	<i>Janetiella oenophila</i>	France, Italy	129
	<i>Lasioptera vitis</i>	E. United States	66
	<i>Dasyneura vitis</i>	E. United States	66
	<i>Schizomyia pomorum</i>	E. United States	66
	Lepidoptera		
	<i>Eupoecila ambiguella</i>	Europe	3, 4, 17, 44, 56, 73
	<i>Lobesia botrana</i>	Europe	78, 91, 92, 96, 101, 112, 133, 145, 153, 165, 175, 183, 188, 197
	<i>Lobesia viteana</i>	E. United States	18, 66, 87, 181, 185
	<i>Argyrotaenia politana</i>	France	32
<i>Argyrotaenia velutinana</i>	E. United States	95, 185	
<i>Platynota stultana</i>	California	7, 8, 115, 173	
<i>Phalaenoides glycine</i>	Australia	18	
<i>Epiphyas postvittana</i>	Australia	18	
<i>Serrodes partitus</i>	S. Africa	132, 199	
Coleoptera			
<i>Craponius inaequalis</i>	E. United States	123	
<i>Lopus sulcatus</i>	France, Italy	32	
Heteroptera			
<i>Euchistus conspersus</i>	California	170	
Chalcidoidea			
<i>Prodecatoma cooki</i>	Florida	2, 55, 164	
Cecidomyiidae			
<i>Contarinia viticola</i>	France	122	
Thysanoptera			
<i>Drepanothrips reuteri</i>	N. America, S. Europe	15, 16, 38-40, 139	
<i>Anaphothrips vitis</i>	Bulgaria, Romania, Greece, Turkey	203	
<i>Haplothrips globiceps</i>	Turkey	54	
<i>Retithrips aegyptiacus</i>	N. Africa, Middle East	152	
<i>Rhipiphorothrips cruentatus</i>	India	10	
<i>Scirtothrips dorsalis</i>	Japan	89	
<i>Heliothrips haemorrhoidalis</i>	World		
<i>Scirtothrips citri</i>	California	170	
Honeydew producers	Homoptera		
<i>Planococcus citri</i>	S. Europe	6, 60	
<i>Planococcus maritimus</i>	California	6, 60	
<i>Planococcus ficus</i>	France	138	

Table 1 (Continued)

Plant organ attacked	Pest		Ref.
	Order	Region	
	<i>Eulecanium corni</i>	S. Europe	20, 32
	<i>Eulecanium persicae</i>	S. Europe	20, 32
	<i>Pulvinaria vitis</i>	S. Europe	20, 32
	<i>Pulvinaria betulae</i>	Romania	127
	<i>Trialeurodes vittatus</i>	California	170
Aerial polyphagous insects	Orthoptera		
	<i>Barbitistes fischeri</i> v. <i>berenguieri</i>	France, Spain, Italy	122
	<i>Ephippiger</i> spp.	S. Europe	122
	<i>Miogryllus convolutus</i>	S. America	113
	<i>Locusta migratoria</i>	S. Europe, N. Africa	122
	<i>Schistocerca peregrina</i>	S. Europe, N. Africa	122
	<i>Doclostaurus maroccanus</i>	S. France	
	Coleoptera		
	<i>Macroductylus subspinosus</i>	E. United States	
	<i>Popilia japonica</i>	California	123
	<i>Anomala</i> spp.	France	85
	Hymenoptera		
	Vespidae	World	156, 195
Polyphagous soil insects	<i>Melolontha melolontha</i>	Europe	85, 121
	<i>Polyphylla fullo</i>	Europe	85, 121
	<i>Anoxia villosa</i>	Europe	85, 121
	<i>Otiorrhynchus sulcatus</i>	Bulgaria	86
	<i>Otiorrhynchus turca</i>	Bulgaria	86
	<i>Agriotes obscurus</i>	Romania	90

One of the areas of penetration into Europe occurred in France in the Gard region (in 1863), where the insect was introduced on some American vinestocks. Westwood recorded it in England in 1867. Little by little the pest spread throughout Europe (186) and then, despite precautions, throughout the whole world. At present, regions that are still untouched are rare. The biology of this species has been studied in each country where it has been introduced. In some places the full cycle of both aboveground and underground stages have been observed, in others only the underground stage is constant. Sometimes rare swarms of winged forms unable to reproduce are found (108, 117, 118, 170, 176). These variations in the insect's life cycle are usually explained by ecological conditions, temperature, and humidity (108, 117, 118). Some have thought also that the isolation of diverse populations of the underground stage and their adaptation to the biotope has permitted the creation of races with particular morphologies or biologies (33, 118, 135). For example, on *Vitis vinifera*, the radicoles produce enormous tuberosities on the roots, thereby causing the death of the rootstock. On American vines they cause only very slight damage because they produce only shallow wounds and because many of them swarm before winter. The origin of this resistance has been the subject of many studies (34-36, 58, 107, 134) because it is the basis for the control of Phylloxera.

The cultivars of *Vitis vinifera* are therefore grafted onto American *Vitis* stocks. Attempts have also been made to hybridize them to avoid grafting, which is a burdensome operation which causes a delay in production. But these resistant directly-producing hybrids, if allowed to spread again to the area of vine cultivation, give a disagreeable taste to the wine. Planting these hybrids has been prohibited in France. Another method of control for radicicole Phylloxera consists of injecting carbon disulfide (CS<sub>2</sub>) into the soil, but this method is very costly and can only be applied over small areas and in vineyards yielding high returns (42). Other methods of chemical control have been recommended for the gall-making stages (12, 42, 98, 177, 179), as well as for the root-feeding stages (98, 99, 175, 178, 187). Note that the radicicoles may also be destroyed by flooding and submersion of the vines for 50 days.

Among the other root-sucking insects one should mention the cicadas, *Cicada plebeja*, *C. orni*, *C. atra* (174), and *Tibicen haematodes* in Europe, (29) and *C. septemdecim* in California (170). The larvae, which live for several years in the soil, pierce the cortical parenchyma and suck the exudate.

Certain coccids are also found on vine roots. The ground mealybug *Rhizoecus falcifer* in California is seldom damaging (170). On the other hand in *Margarodes meridionalis* in California (170), *Eurhizococcus brasiliensis* in Brazil (72), *M. vitis* Chile, and *M. capensis* and *M. greeni* in South Africa (174), the globular females fix themselves on the rootlets and thus weaken the stocks.

Coleopterous larvae, which have a subterranean life, cause damage by gnawing the roots. In Europe *Bromius obscurus*, which was previously a pest of some importance, has now become quite rare. The larvae destroy the cortical part and the superficial wood of the roots (19). They live with difficulty on the roots of American *Vitis*, which explains their rarity after the widespread use of American rootstocks following the crisis caused by Phylloxera. The adult bites and pierces the parenchyma of the leaves and the epidermis of the young grapes following a sinuous line, hence the name *writer* (*écrivain*) which has been given to it. The species is parthenogenetic and only the female is known. It has been introduced into North America, where another species, the grape rootworm beetle, *Fidia viticida*, causes similar damage (66). In India a third Eumolpina, *Scelodonta strigicollis*, behaves in the same way as the two above-named species. The wounding caused by the larvae on the roots often leads to the death of the rootstock. The adult gnaws the surface of the young leaves of the vine shoots and the young grapes (109).

Other oligophagous Coleoptera larvae are also injurious to the roots of the vine. Around the Mediterranean the genus *Vesperus*, with *V. xatarti*, *V. strepens*, and *V. luridus*, causes local but often serious damage (63, 146); in addition, *Pentodon punctatus*, *P. idiota*, *P. bispinosus* (19, 32, 122), *Phyllognathus excavatus* (85), and a Tenebrionidae, *Opatrum sabulosum* (32), have larvae which attack the collar of the young rootstocks and gnaw the graft calluses.

The caterpillar of *Vitacea polistiformis* also causes considerable damage to roots in the Missouri region, where certain vineyards have been completely destroyed (59). Their life cycle lasts three years. The first-instar larvae have very poor resistance to drought (155).

*Wood-Damaging Pests*

**ON THE TRUNK** Termites attack rootstocks that are in poor condition for any of several reasons such as lesions caused by frosting, by agricultural implements or poor circulation of the sap, due, for example, to an incorrect choice of a graft stock that is ill-adapted to the soil. Examples in France are the yellownecked termite *Calotermes flavicollis* and, more rarely, *Reticulitermes lucifugus* (71), and in California *Reticulitermes hesperus* (170).

*Cossus cossus* (70), whose large, red caterpillar sometimes destroys the rootstock in Europe, has its homolog in *Paropta paradoxus* (141) in Israel and in Egypt. Finally, in Bulgaria a cerambycidae, *Anaglyptus mysticus*, has recently been observed (163). The larvae destroy the cambial tissue, causing the death of 13% to 15% of the rootstock of four- to seven-year-old vines.

The control of trunk-attacking species is difficult. For the termites and *Cossus*, the only method of control is to maintain the vine in excellent condition. On the other hand, it seems that oleoparathion preparations or dichlorvos are effective for controlling *A. mysticus* (163).

**ON THE SHOOTS** *Scale insects* In Europe and America, two Diaspididae, *Eulecanium corni* and *Pulvinaria vitis*, attack the shoots, as does the grape scale *Diaspidiotus uvae* in California (170). They are damaging not only because they remove the sap, but also because of the toxicity of their saliva. Other species of Coccidae [*Planococcus citri* (20, 32) and *Planococcus ficus* (138) in Europe and *Planococcus maritimus* in America (170)] cause a similar nuisance, but they are mobile and damage principally the young grapes because of the coccids production of honeydew and the subsequent growth of the sooty mould fungi on the honeydew.

**Others** Other insects attack the vine shoots but the damage inflicted is never very important, as in the following examples:

1. Cicadas make fusiform egg-laying cicatrices.
2. *Ceresa bubalus* has rings of egg-laying punctures around the shoot which destroy the vascular system and produce a spectacular callus above the wound (168, 192). This species passes part of its life cycle on fruit trees and the vine and then part on herbaceous cultivated or adventitious plants. An attempt at biological control using the egg parasite *Polynema striaticorne* is underway (143).
3. The Bostrychids—*Apate sexdentatum* (129), *Apate muricata*, *Schistoceros bimaculatus*—attack decaying stocks and penetrate at bud level to lay eggs there. The larvae bore longitudinal galleries in the pith (122).
4. The branch and twig borer *Polycaon confertus* is found in California. The adult feeds by chewing a hole in the base of the young shoots, which then may be broken by the wind. The larva lives in the main branches of the stock in decaying or dead wood (170).
5. The vine sawfly, *Macrophya strigosa*, lays its eggs in the green shoots; the larva descends into the pith and bores its gallery until it reaches the buds. The damage can be locally important (122).
6. Finally, numerous Hymenoptera species make notches in the pith to nest there.



### Bud Pests

The damage caused by these insects always has a catastrophic effect on the crop. In Europe, the tiger moth, *Arctia caja*, or *chenille bourrue*, so named because of its hairiness, overwinters as a caterpillar on weeds and afterwards passes onto the vine devouring five to six buds a day at the time of bud break and later on attacks the young shoots and the floral buds. A method of chemical control has been perfected (27); in addition, an entomogenous fungus, *Entomophthora sphaeroderme*, produces fierce and deadly epidemics among the caterpillar populations.

Throughout the world numerous species of noctuids (cutworms) are known as bud destroyers. In Europe the most important pests are *Autographa gamma*, *Agrotis segetum*, *Euxoa nigricans*, *Triphaena pronuba*, etc (53). In America some of the most prominent pests are the variegated cutworm *Peridromia margaritosa*, the Greasy cutworm *Agrotis ipsilon* (170), and the cutworms, *Anagrotis barnesis* (134) and *Rhynchagrotis cupida* (151). The caterpillars of all these species pass the day a few centimeters below the soil at the foot of the rootstocks; they come out at night to feed on the buds. Although the viticulturist was for a long time defenseless against them, they can now be effectively controlled (26, 53).

Some Coleoptera in the adult stage may also attack buds, e.g. in Europe the vine grubs (*coupe bourgeons*), *Peritelus sphaeroides* and *P. noxius* (84), and in America the diurnal click beetle *Limonium canus*, the grape bud beetle, *Glyptoscelis squamulata*, whose adults penetrate into the buds without damaging the scales, and the little bear beetle, *Pocalta ursina*, which does only occasional damage (170). In South Africa three species of vine grub are found: *Phlyctinus callosus*, *Eremnus cerealis*, and *E. stulosus* (24, 25).

Finally, one must mention the mite, *Calepitrimerus vitis*, which hinders the development of the buds and young shoots by the damage caused by infestations. The internodes are short and sometimes the grapes abort (120, 148). *Eriophyes oculivitis* (11) in Egypt and *E. vitineusgemma* (119) in Moldavia have recently been described, and their damage is similar to that of the preceding species. Some tydeids are effective predators of these phytophages (157, 159, 202).

### Leaf Pests

**LEPIDOPTERA** The pyralid *Sparganothis pilleriana*, once common throughout all French vineyards, appears to have declined sharply (169). It has only one generation per year, and the first-instar caterpillar, which hibernates in a cocoon under the bark of the vinestock, can be destroyed at that moment by sprayings of sodium arsenite. Use of this chemical is currently allowed, but it is being replaced by phosphoric acid esters (145). At the three-to-four-leaf stage the larvae are killed by toxaphene, which is also active against the hairy caterpillar (*chenille bourrue*) (149). Numerous indigenous parasites control the populations of the pyralid. The caterpillars have a marked preference for the plants whose leaves are rich in protein (142). Among the sphingid caterpillars, the whitelined sphinx moth, *Celerio lineata*, is the most damaging in France as well as in California. In the latter region, the achemon sphinx moth, *Pholus achemon*, is also found (170). The larvae of *Antispila rivillei* have been recorded as vine leafminers in Georgia (57).

Another Lepidoptera, the western grapeleaf skeletonizer, *Harrisina brillans*, which has passed from Mexico into California, has curious, gregarious caterpillars with black-and-red-striped, yellow bodies. The larvae destroy the leaf blade without damaging the upper epidermis (140, 170). There have been attempts at microbiological control (123) and biological control (84a). In India the caterpillars of *Sylepta lunalis* destroy just as completely the parenchyma of the leaves which they roll (136).

**COLEOPTERA** The halticids, *Haltica lythri* ssp. *ampelophaga* in Europe and *Haltica chalybea* and the grape flea beetle *Haltica torquata* (170) in California, have gregarious larvae which also eat the leaf blade without damaging the cuticle.

The leafrollers (*cigariers*), *Byctiscus betulae*, in France and the grape leafroller, *Desmia funeralis*, in America have females which cut the leaves and roll them into "cigars" within which they lay their eggs and which serve as food for the larvae. Treatment trials against this last species have shown that a preparation based on *Bacillus thuringiensis* is as effective as some chemical pesticides (carbaryl) (5, 93). Attempts at biological control by the rearing and release of natural parasites have given encouraging results (69).

**HEMIPTERA** Only one species of aphid has been recorded on the vine, *Aphis illinoisensis*. It produces heavy infestations on young shoots and on the leaves in summer to the east of the Mississippi River. It is dioecious and migrates in autumn onto its principal host (123).

On the other hand, a number of cicadellid species are vectors of phytopathogenic agents. Pierce's disease, found in North America, has 20 species as vectors (83, 144, 170, 200). One of the most efficient among these is *Philaenus spumarius*, a cercopid, which also occurs in France where very fortunately Pierce's disease is not found. The golden yellows, a disease which appeared about 1950 in the southwest of France, is also caused by a molicute (79). The vector is *Scaphoideus littoralis*, a species of American origin. The disease has spread and at present affects Switzerland, Italy, Sardinia, Corsica, and perhaps Germany (45, 50, 51, 76). Furthermore, the disease of black wood (*bois noir*), which is rife in Burgundy and in the Jura, very probably has a cicadellid as vector also (49, 194).

The cicadellids can also cause direct damage by piercing the phloem into which their toxic saliva is injected (30, 31, 160, 162, 189, 190), e.g. *Empoasca flavescens*, which has multiplied in France during the last few years, perhaps because of the physiological condition of the vine after pesticide treatment (57). It causes a crinkling of the leaf, a hardening of the leaf blade, and a browning of the nerves (14). The leaves dry up from the periphery towards the center and then fall (131, 193). Quite often the treatment against the grape moths (*tordeuses*) is sufficient to control *Empoasca vitis* (= *E. flavescens*) (13, 52, 128). *Empoasca libyca* (191) causes the same damage in hotter regions, such as southern Spain, Sicily, Sardinia, North Africa, Israel, and Tanganyika. *E. decipiens*, though not a pest in France, is one in Baluchistan. Recently, two Flatidae, *Flata ferrugata* and *Unnata intracta*, have infested the vine in the Punjab (171). Finally, other species—*Zygina rhamni* in

France, *Erythroneura adanae vitisuga* in Bulgaria (102), *E. comes*, *E. variabilis*, and the grape leafhopper, *E. elegantulae*, in California (96, 173), and *E. ziczac* in British Columbia (124)—are also damaging to the vine. *Ceresa bubalus*, already mentioned for the damage it does on the shoots, also causes a crinkling and a reddening of the leaves.

Other Hemiptera also damage the leaves, such as *Nyzius senecionis*, which pierces at bud level and whose toxic saliva produces the drying-up of what is above the wound. The false chinch bug *Nysius ericae* lives in Europe and America where it causes damage similar to that of the above-named species (22). These insects migrate by walking from their plant host to the vine. They can thus be controlled by creating a barrier consisting of a ten-meter band of land powdered with lindane.

**ACARINES** The punctures of the cell feeders gravely damage the leaves, which dry up and fall. The cell feeders in Europe consist of two Tetranychidae: the yellow spider mite, *Eotetranychus carpini*, which lives on the underneath side of the leaves and whose punctures cause red or yellow stains, according to the vine cultivar, along the nerves (65, 81), and the red spider mite, *Panonychus ulmi*, which lives on both sides of the leaves, causing them to take on a grayish tint (47). Recently, heavy infestations of *Tetranychus atlanticus* have appeared in the vineyards of Mediterranean Languedoc, as a consequence of using herbicides (A. Rambier, 1975, personal communication). Other, less-damaging species exist on the vine in France and throughout Europe (21, 126). In California *Tetranychus pacificus* is the homolog of the French tetranychids on the vine (104, 105, 111, 173), and *Tetranychus flavus* dries up the buds and the young, unopened leaves. Their most effective predator is *Metaseiulus occidentalis* (9, 104, 105). *Oligonychus mangiferae* has recently been observed in India, where it produces damage similar to that from the tetranychids on the French vines (167). The mite problem did not exist before the application of synthetic insecticides against the grape moths (*tordeuses*). Many authors have tried to determine the causes inducing these multiplications (45, 56, 158, 173). They are multiple, principally including the selection of resistant races, the suppression of predators (typhlodromids, anthocorids, thrips, etc) (37, 65, 147, 157), and the effects of insecticides on the physiology of the vine and on the mites are in a sense favorable to the fecundity and longevity of the latter (56). Because many synthetic insecticides induce heavy mite infestations, the control of other insects damaging to the vine is often rendered more difficult (3, 17, 73).

**GALLMAKERS** Leaf blister is caused by the mite *Eriophyes vitis*. The gall occurs as a depression on the underneath part of the leaf. It is lined with white hairs, within which are found the minute mites (77, 126, 148). *E. vitigenus gemma* and *E. oculivitis*, both recently described [the first is from Moldavia (119), the second from Egypt (11, 202)], do not form a blister but instead dry up the buds.

Some other species can also be gallmakers. By its punctures the gall-living form of *Phylloxera* provokes the formation of galls on American vines and on certain Franco-American hybrids. The gallforming female lives in a globular gall, which opens onto the upper surface of the leaf. Only the young larvae can leave by passing

through the hairs that obstruct the orifice. The sensitivity of different vine cultivars to attack by the gallforming aphids has been tested (180), and diverse insecticides have tried as controls against them (179).

The leaf cecidomyid *Janetiella oenophila* forms lenticular galls between the two cuticles of the leaf (129). One can mention also in America the grapevine tomato gallmakers, *Lasioptera vitis* and *Dasyneura vitis*, as well as another Diptera, *Schizomyia pomorum* (66).

### *Fruit Pests*

The grape moths (*tordeuse de la grappe*) give the most trouble to French viticulturists. The grape tortrix *Lobesia botrana* and the grape tineid *Eupoecilia ambiguella* are often confused by the experts because they have fairly similar biologies and synchronous flight periods, and they cause nearly the same damage. In France *L. botrana* has three annual generations and lives principally in dry situations, whereas *E. ambiguella* has only two generations and lives in humid situations. Their biology (44, 92, 112, 133, 165) and methods of control have been the object of many studies in all countries (3, 4, 68, 91, 97, 101, 112, 145, 153). The treatments used are of two types: preventive, by applications before hatching of the eggs, or curative, by using control measures against the young caterpillars. The treatments are applied after announcements are broadcasted by the Plant Protection Service. In effect, the biology of the two species depends directly on meteorological conditions (175), and it is necessary to have recourse to trapping to determine with precision the date and importance of the flight of adult moths. The captures from food traps are often difficult to identify (183, 197), and confusions with non-pest species can occur (96). The latter can be avoided by the use of sexual traps, which moreover have a high efficiency (78, 188). Observation of egg laying in the vines is being used more and more to fix with certainty the necessity and date of treatment. The damage by the second and especially that done by the third generation of *Eupoecilia* is often severe. The caterpillars penetrate into the grapes, and the entry holes favor the establishment of the fungus of the gray mold *Botrytis cinerea* (3). It is noteworthy that the treatments against the grape moths, in particular those with phosphoric esters, have induced heavy infestations of mites in the vineyards (3, 17, 56, 73).

In the eastern United States, the grape berry moth, *Lobesia viteana*, has the same biology and causes the same damage as the French *Eudemis*, but because pupation occurs not under the bark of the vine stock but instead in a rolled-up leaf on the ground, efficient control by cultural means (66, 87, 181) and by using *Bacillus thuringiensis* (28) is possible. A confusion operation with the aid of sex pheromones has also given good results (185).

The small grape moth *Argyrotaenia politana* was discovered for the first time in Montpellier, France, in 1954. It is questionable whether it is a species newly adapted to the vine, coming from the numerous apple orchards planted a few years before in the region, or whether it has been confused with *Eudemis*, which the adult closely resembles. The damage inflicted by the two species is the same, only the treatment dates differ (32). In the eastern half of the United States, a related species, the redbanded leafroller, *Argyrotaenia velutinana* occurs (95, 185). In California

*Platynota stultana* (7, 8) has become an important pest during the last few years (115, 173); all of the nonligneous parts are attacked, but the greatest damage is to the grapes. The numerous wounds inflicted on the epidermis permit fungal spores to penetrate and cause rot. Crop losses can reach 15%. In Australia the vine moth, *Phalaenoides glycine*, and the light brown apple moth, *Epiphyas postvittana*, cause the same damage and are justifiably controlled by the same methods as the above-named species (18). A Coleoptera, the grape curculionid *Craponius inaequalis* lays its eggs in the grapes, where the larvae consume the seeds and the pulp (123). In California the consperse stink bug, *Euchistus conspersus*, which habitually lives on low plants, pierces the grapes and thereby greatly depreciates the crop of table grapes (170). A chalcid in Florida, *Prodecatoma cooki*, whose larva lives in the pulp and the seeds (2, 55, 164) and the fruit-piercing moth, *Serrodus partitus* (F.), in South Africa must also be noted (132, 199). Finally, two species whose very infrequent damage can be considered as negligible in France must be mentioned: the polyphagous capsid *Lopus sulcatus*, which sometimes passes onto the vine (32) and whose larvae and adults pierce the flower buds that blacken and fall, and the vine cecidomyid *Contarinia viticola*, cause similar damage (122).

Many species of thrips (Thysanoptera) attack the vine in the world. They are very rarely mentioned because on account of their small size they pass unnoticed and their damage is attributed to other pests. The grape thrips, *Drepanothrips reuteri*, probably originated from California where its biology and control have been studied (15). Introduced into Europe, it had been recorded as damaging solely American vines (139). During the last ten years it has been recorded as damaging hybrid vines first of all (38), then French vines (16, 39, 40). Over and above the damage it causes to the leaves (necrosis or holes in the blade), the punctures of the larvae and of the adults produce a toxic reaction which retards the development of the shoots and causes a certain amount of abortion at the flowering period. The most severe damage is recorded on the grapes where it causes necrosis and suberization of the epidermis; this considerably decreases the varieties of table grape. Its biology and methods of control have been studied in Europe (16, 38–40). The treatments against the second generation of the grape moths are also valuable against thrips. Some vicariant species in other climates occupy the same ecological niche as this species and cause similar damage to the grapes. These include, for instance, *Anaphothrips vitis* (203) in Bulgaria, Romania, and Greece; *Haplothrips globiceps* (54) in Turkey; *Retithrips aegyptiacus* in the Middle East (152), Egypt, and North Africa; *Rhipiphorothrips cruentatus* (10) in India; *Scirtothrips dorsalis* in Japan (89); and *Heliothrips haemorrhoidalis*, a polyphagous species of warm and warm temperate climates which also passes onto the vine in Chile, for example. Other species can also occasionally attack the vine: *Scirtothrips citri* (170) close to citrus orchards and *Hercothrips fasciatus*, as well as the grass thrips *Frankliniella moultoni*, *F. occidentalis*, and *F. minuta* in the United States.

### *Honeydew Producers*

Certain insects are also a nuisance indirectly because of the production of a sweet honeydew. This secretion serves as a substrate for a black fungal growth, the sooty mould, which greatly depreciates the quality of table grapes.

Among the coccids, the females of *Planococcus citri*, mobile in all stages, have soft integument covered with a white bloom. The species can be effectively controlled by releases of a coccinellid, *Cryptolaemus montrouzieri* (13). *Pseudococcus maritimus* (6, 60) is the California homolog of the above species. One can also mention *Pulvinaria vitis*, *Eulecanium corni*, and *Eulecanium persicae*, which are controlled successfully by the entomogenous fungus *Beauveria bassiana*, and *Pulvinaria betulae* in Romania (127). These species are quite often controlled by the treatments against the grape moths. Furthermore, since 1969, perhaps because of the employment of organophosphorus insecticides, a new species that is locally very damaging has multiplied throughout the Mediterranean basin: *Planococcus ficus* (138). In California the aleurodid *Trialeurodes vittatus* also produces honeydew.

### *Polyphagous Aerial Insects*

These are principally the Orthoptera. Among the Ensifera, the *Boudrague*, *Barbitistes fischeri* var. *berenguieri* (122) and three species of the genus *Ephippiger*—*E. ephippiger*, *E. bitterensis*, and *E. terrestris*—gnaw the foliage and the green or ripe grapes in the Mediterranean vineyards. The damage, although episodic, can be important (122). Among the acridians, *Locusta migratoria*, *Schistocerca peregrina*, and *Dociostaurus maroccanus* have gregarious bands that are particularly destructive. *Miogryllus convolutus* has recently been recorded as damaging vines in South America (113).

Some polyphagous Coleoptera, in particular the scarabs, can sometimes destroy the green parts of the vine. In the eastern United States the rose chafer, *Macrodactylus subspinosus*, and the Japanese beetle *Popilia japonica*, whose larvae also attack the roots (123), are found. In France there are the green vine chafer, which includes in reality a complex of three related species—*Anomala vitis*, *A. dubia*, and *A. ausonia* (85)—which are found only in the vines on the sands of the Mediterranean coast and whose larvae only feed on decomposing plant material. Finally, practically everywhere in the world (156, 164, 195) the wasps (Vespidae) attack the ripe grapes, causing serious damage to table varieties.

### *Polyphagous Soil Insects*

Mostly polyphagous soil insects are cockchafer grubs, larvae of various species: the common cockchafer, *Melolontha melolontha*, the pine chafer, *Polyphylla fullo*, and the hairy chafer, *Anoxia villosa* (85, 121). In Bulgaria (86) *Otiorrhynchus sulcatus* and *O. turca* larvae are recorded damaging the roots. Larvae of *Agriotes obscurus* cause damage in Romania (90).

## CONCLUSION

In France, the major arthropod grape pests are the grape leaf rollers, *Lobesia botrana* and *Eupoecilia ambiguella*, and the tetranichids, *Eotetranychus carpini* and *Panonychus ulmi*. The two mite species were not pests until 1950 when the ill-considered use of organophosphates in controlling leaf rollers caused catastrophic outbreaks of mites. To avoid such outbreaks, studies were undertaken to promote the concept of integrated control in vineyards (1, 73, 148).

To control leaf rollers knowledgeably, levels of economic tolerance were established; these vary in relation to the phenologic stages of the vine, the climate, and the value of the crop. Population dynamics were based on data from traps, by using light, attractants, and sex pheromones, as well as from observing egg laying by adult moths. The hibernating forms of mites were carefully counted to predict future outbreaks.

French workers have shown that the use of certain pesticides causes mite outbreaks in two ways. First, pesticides destroy mites' natural predators, *Scolothrips* spp., *Orius vicinus* (Anthocoridae), *Stethorus punctillum* (Scimniini), and *Typhlodromus* spp. (147, 148). Additionally, certain insecticides and fungicides alter plant metabolism to produce conditions more favorable to mite growth and reproduction. In this way it has been shown that soluble nitrogen (amino acids) and reducing sugars play a large role in the nutrition and reproduction of mites (1, 56, 57). Outbreaks of leafhoppers (*Empoasca flavescens*) analogous process. For this reason, after detailed investigations, a list of pesticides and their effects on pests and auxiliaries was compiled (1). The problem of tetranychid outbreaks on grapes has also been studied in California (73). Ecological studies have made it possible to intensify the predation of *Metaseiulus occidentalis*, which attacks *Eotetranychus willamettei* (a minor pest) when *Tetranychus pacificus* is absent. As in France, pesticides that spare predators were selected.

To avoid the disadvantages of chemical control, recent experimental work has tested the effectiveness of *Bacillus thuringiensis* against larvae of *Lobesia botrana* (153). These are very susceptible but two difficulties are not yet resolved. The fact that eggs are laid and hatched successively over protracted periods and the behavior of the insect make contact between larvae and spores uncertain. Preventive treatment therefore cannot be based on defined infection thresholds.

Other projects, using entomophagous insects, have also been initiated. Two parasites of *Harrisina brillians*, the tachnid *Sturmia harrisinae*, and the hymenopteran *Apanteles harrisinae*, as well as a virus introduced from Arizona, were effective in lowering *H. brillians* populations in San Diego County, California.

Controlling the leafhopper *Erythroneura elegantula* by the mymarid *Anagrus epos* has been improved by providing refuges of *Rubus* sp. that harbor the nonpest species *Dikrella cruentata*, the eggs of which allow hibernation of the parasite. Other trials of entomophagous insects, e.g. against *Desmia funeralis*, are in progress.

There is no question that the continuing progress of ecological sciences will lead to a greater effectiveness of integrated control programs for grape pests as they have for other crops.

## Literature Cited

1. A C T A. 1975. Lutte intégrée en vignoble. *Assoc. Coord. Tech. Agric. Spéc.* 5: 62 pp.
2. Adlerz, W. C. 1972. *Prodecatoma cooki* (How.), a seed Chalcid on Florida grapes. *J. Econ. Entomol.* 65(5):1530
3. Agulhon, R. 1973. Trois problèmes phytosanitaires étudiés dans le vignoble en 1972: l'excoriose, les Tordeuses de la grappe, les acariens. *Vignes et Vins Mai 1973*, 219:15-22
4. Alexandri, A. A. 1973. Efficacy of some insecticides in the control of the grape moth (*Polychrosis botrana* Schiff.) (Rouman). *Ann. Inst. Cerc. Pentru. Protect. Plant.* 9:507-14
5. Aliniaze, M. T., Jensen, F. L. 1973. Microbial control of the grape leaf folder with different formulations of *Bacillus thuringiensis*. *J. Econ. Entomol.* 65(1):157-58
6. Aliniaze, M. T., Stafford, E. M. 1972. Control of the grape mealybug on "Thompson Seedless" grapes in California. *J. Econ. Entomol.* 65(6):1744
7. Aliniaze, M. T., Stafford, E. M. 1972. Notes on biology, ecology and damage of *Platynota stultana* WLSM on grapes. *J. Econ. Entomol.* 65(4):1042-44
8. Aliniaze, M. T., Stafford, E. M. 1973. Management of grape pests in Central California vineyards. 1. Cultural and chemical control of *Platynota stultana* on grapes. *J. Econ. Entomol.* 66:154-57
9. Aliniaze, M. T., Stafford, E. M., Kido, H. 1974. Management of grape pests in Central California vineyards: toxicity of some commonly used chemicals to *Tetranychus pacificus* and its predator *Metaseiulus occidentalis*. *J. Econ. Entomol.* 67(4):543-47
10. Ananthkrishnan, T. N. 1971. Thrips in Agriculture, Horticulture and Forestry- Diagnosis, Bionomics and Control. *J. Sci. Indust. Res.* 30(3):113-46
11. Attiah, H. H. 1969. *Eriophyes oculivitis* n. sp. a new bud mite infesting grapes in the U.A.R. *Bull. Soc. Entomol. Égypte* 51:17-19
12. Avdyshev, Sh. E. 1971. Prospects for chemical control of grape vine Phylloxera. *C.R. Int. Congr. Moscou, 13th 1968*:206-7
13. Aykac, M. K., Erguder, T. M. 1974. A study of control measures against *Planococcus citri* (RISSO) in the vineyards of Tokat province. *Sams. Böl. Zir. Mucad. Arast. Enstit.* 43:171-72
14. Baggjolini, M., Canevascini, V., Caccia, R. 1972. La Ciccadelle verte (*Empoasca flavescens* F.) cause d'importants rougissements du feuillage de la vigne. *Bull. OEPP* 3:43-49
15. Bailey, S. F. 1942. The grape or vine thrips *Drepanothrips reuteri* Uzel. *J. Econ. Entomol.* 35(3):382-86
16. Baillod, M. 1974. Dégâts de thrips sur vigne en Suisse romande. *Rev. Suisse Vitic. Arboric. Hortic.* 6(2):45-48
17. Baillod, M. 1974. La protection de la vigne contre l'araignée rouge *Panonychus ulmi* (Koch) et l'araignée jaune commune *Tetranychus urticae* (Koch). *Rev. Suisse Vitic. Arboric. hortic.* 6(1): 17-22
18. Baker, B. T. 1974. Practical pest and disease management in vineyard. *Aust. Grapgrower Winemaker April* pp. 1-10
19. Balachowsky, A. S. 1962-1972. *Entomologie Appliquée à l'agriculture*. Vol. 4 Paris:Masson
20. Balachowsky, A. S., Mesnil, L. 1935. *Les insectes nuisibles aux plantes cultivées*, Vol. 1-2. Paris:Masson. 1921 pp.
21. Balevski, A., Martinov, S., Nachev, P. 1970. The grape-vine mite (*Brevipalpus lewisi* (McGr.) the cause of dessication of grape vines in the Vidin district. *Gradinar. Lozar. Nauka*, 7(6):67-76
22. Barnes, M. M. 1970. Grape pests in Southern California. *Calif. Agric. Exp. Stn. Circ.* 553:1-10
23. Beffa, D. G. 1949. *Gli Insetti Dannosi All'agricoltura e i Moderni Metodi e Mezzi di Lotta*, ed. Miland: Hoepli. 978 pp.
24. Berg, H. C. Van den. 1971. The biology and control of vine snout beetle. *Decid. Fruit Grow.* 21:83-85
25. Berg, H. C. Van den, Giliomee, J. H. 1972. Aspects of the ecology and behaviour of *Eremnus cerealis* Marshall. I. The emergence and distribution of the adults and the distribution of the larvae in the vineyard. *J. Entomol. Soc. South. Afr.* 35(1):171-76
26. Berville, P. 1954. La lutte contre les vers gris de la vigne. *Phytoma* 56:26
27. Berville, P., Terral, A. 1958. Observations et essais de traitement sur les Chenilles bourrées. *Phytoma* 102:7-10
28. Biever, K. D., Hostetter, D. L. 1975. *Bacillus thuringiensis* against lepidopterous pests of vine grapes in Missouri. *J. Econ. Entomol.* 68(1):66-70
29. Blunck, H. 1956. See Ref. 172, pp. 190-95



30. Boller, E. 1971. Untersuchungen an der Rebzikade (*Empoasca flavescens* F.) und am embindigen Traubenwickler (*Clysia ambiguella* Hb.) in der Ostschweiz. *Schweiz. Z. Obst Weinbau* 106:651-60
31. Bonfils, J., Leclant, F. 1972. Reconnaissance et nuisibilité des Cicadelles sur la vigne. *Prog. Agric. Vitic.* 89(14):343-55
32. Bonnemaïson, L. 1962. Les ennemis animaux des plantes cultivées et des forêts, Vol. 3 Paris:SEP. 1502 pp.
33. Börner, C. 1925. Die neuen Forschungen zur Reblausrassefrage. *Dtsch. Weinbau* 1:5-8; 2:18-19; 3:29; 4:36-37; 5:48-49; 6:118-124
34. Börner, C. 1939. Anfalligkeit, Resistenz und Immunität der Reben gegen die Reblaus. *Z. Hyg. Zool. Schädlinge be Kämpf.* 31:274
35. Boubals, D. 1966. Etude de la distribution et des causes de la résistance au Phylloxera radicolle chez les Vitacées. *Ann. Amélior. Plant.* 16(2):145-84
36. Boubals, D. 1966. Hérité de la résistance au Phylloxera radicolle chez la vigne. *Ann. Amélior. Plant.* 16(4):327-47
37. Bournier, A. 1954. A propos des thrips de la vigne. *Prog. Agric. Vitic.* 142:104
38. Bournier, A. 1957. Le thrips de la vigne: *Drepanothrips reuteri*. *Ann. Ecole Natl. Agric. Montpellier* 30(1):145-57
39. Bournier, A. 1962. Dégâts de thrips sur vignes françaises. *Prog. Agric. Vitic.* 79(7):164-73
40. Bournier, A. 1965. Sur l'adaptation d'un Insecte nuisible à une culture *Drepanothrips reuteri* Uzel sur vignes françaises. *C.R. Congr. Int. Entomol. Londres 12th, 1964. Son 9a, Agric. Entomol.* p. 537
41. Bovey, R. 1967. *La Défense des plantes Cultivées*, 176-218. Lausanne: Payot. 847 pp.
42. Branás, J. 1968. Le Phylloxera et les Insecticides. *Prog. Agric. Vitic.* 85(17):401-9
43. Branás, J. 1974. *Viticulture*. Montpellier: ENSA. 990 pp.
44. Carbo Sagner, J., Ripolles Moles, J. L., Bricio Sanz, M., Fabregas Sole, C. 1973. Studies on the bionomics of the vine moths (*Lobesia botrana* and *Clysia ambiguella*). *Bol. Inf. Plag.* 112:49-51
45. Carle, P. 1965. Essais de pesticides en plein champ contre *Scaphoideus littoralis*. *Phytiatr. Phytopharm.* 14:29-38
46. Carles, P., Chaboussou, F., Harry, P. 1972. Influence de la nature du porte greffe de la vigne sur la multiplication de l'araignée rouge *Panonychus ulmi* Koch aux dépens d'un même greffon: le Merlot rouge. *C. R. Seances Acad. Agric. Fr.* 58(17):1403-15
47. Carmona, M. M. 1973. The presence of the mite *Panonychus ulmi* KOCH on vines. *Agricoltura* 4:16-21
48. Castro, A. R. 1965. Plagas y enfermedades de la vid. *Inst. Nac. Invest. Agric.* p. 757
49. Caudwell, A. 1961. Etude sur la maladie du bois noir de la vigne: ses rapports avec la flavescence dorée. *Ann. Epiph.* 12(3):241-68
50. Caudwell, A., Moutous, G., Brun, P., Larrue, J., Fos, A., Blancon, G., Schick, J. P. 1974. Les épidémies de flavescence dorée en Armagnac et en Corse et les nouvelles perspectives de lutte contre le vecteur par des traitements ovicides d'hiver. *Bull. Tech. Inf. Minist. Agric.* 294:1-12
51. Caudwell, A., Ottenwaelter, M. 1957. Deux années d'études sur la flavescence dorée nouvelle maladie grave de la Vigne. *Ann. Amélior. Plant.* 4:359-93
52. Caudwell, A., Schwester, D., Moutous, G. 1972. Variété des dégâts des Cicadelles nuisibles à la vigne. Les méthodes de lutte. *Prog. Agric. Vitic.* 89(24):583-90
53. Cayrol, R. A. 1972. Famille des *Nocuidae*. See Ref. 19, pp. 1255-520
54. Cengiz, F. 1973. Recherches sur les Thysanoptères vivant sur vigne dans les territoires d'Izmir et de Mamisa: leurs caractères morphologiques, les plantes hôtes, leurs dégâts et leurs ennemis naturels. *Thèses Doc. Etat Bornova Izmir.* 112 pp.
55. Cermeli, L. M. 1973. The wasp attacking grapes (*Vitis vinifera*) a new pest of this crop in Venezuela. *Agron. Tropic.* 23(4):413-17
56. Chaboussou, F. 1969. Recherches sur les facteurs de la pullulation des Acariens phytophages de la vigne à la suite des traitements pesticides du feuillage. *Thèse Doc. Sci. Paris.* 238 pp.
57. Chaboussou, F. 1971. Le conditionnement physiologique de la vigne et la multiplication des Cicadelles. *Rev. Zool. Agric. Pathol. Vég.* 70(3):57-66
58. Chebotar, T. I. 1971. See Ref. 201, pp. 96-102
59. Clark, G. N., Enns, V. R. 1964. Life history studies of the grape root borer in Missouri. *J. Kans. Entomol. Soc.* 37(1):56-63
60. Cone, W. W. 1971. Grape mealybug control in Concord grape field trials in

- central Washington. *J. Econ. Entomol.* 64(6):1552-53
61. Couturier, A. 1938. Remarques sur la biologie de *Ceresa bubalus* F. membracide d'origine américaine. *Rev. Zool. Agric.* 37(10):145-57
  62. Delassus, M., Lepigre, A., Pasquier, R. 1930. Les ennemis de la vigne en Algérie et les moyens pratiques de les combattre. I. Les parasites animaux. Alger: Carboneil. 249 pp.
  63. Delmas, H. G. 1954. Le vespère de la vigne. Techniques de lutte. *Rev. Zool. Agric. Appl.* 53:110-20
  64. Delmas, R. 1956. Lutte contre les ennemis animaux de la vigne. *C.R. Cong. Int. Vigne Vin*, 8th, pp. 57-71
  65. Delmas, R., Rambier, A. 1954. L'invasion des araignées rouges sur la vigne. *Prog. Agric. vitic.* 142:34-35; 101-4
  66. Demaree, J. B., Still, G. W. 1951. Control of grape diseases and insects in eastern United States. *Farm. Bull.* 1893: 36 pp.
  67. Demetrashvili, M. I. 1971. The grape mining. *Zashch. Rast.* 16(8):53
  68. Dirimanov, M., Kharizanov, A. 1972. Tests of preparations for the control of the larvae of variegated grape moth. *Zashch. Rast.* 20(10):23-26
  - 68a. Douth, R. L., Nakata, J., Skinner, F. E. 1966. Dispersal of grape leafhopper parasites from a blackberry refuge. *Calif. Agric.* 20:14-15
  69. Douth, R. L., Nakata, J., Skinner, F. E. 1969. Parasites for control of the grape leaf folder. *Calif. Agric.* 23(4):4
  70. Feron, J., Audemard, H., Balachowsky, A. S. 1966. Superfamille des *Cossoidea*. See Ref. 19, 1:39-59
  71. Ferrero, F. 1973. Les Dégâts de termites dans le cru de Banyuls. *Phytoma* 25(251):25-27
  72. Figueiredo, E. R. de 1970. Nova praga da videira em Sao Paulo *Eurhizococcus braziliensis* (Hempel). *Biologico* 36(9): 229-34
  73. Flaherty, D. L., Lynn, C. D., Jensen, F. L., Luvisi, D. A., 1969. Ecology and integrated control of spider mites in San Joaquin vineyards. *Calif. Agric.* 23(4):11
  74. Galet, P. 1968. *Précis d'Ampelographie Pratique*. Montpellier:ENSA. 230 pp.
  75. Galet, P. 1970. *Précis de Viticulture*. Montpellier:ENSA. 490 pp.
  76. Gartel, W. 1965. Untersuchungen über das Auftreten und das Verhalten der "Flavescence dorée" in den Weinbaugebieten an Mosel und Rhein. *Weinbau Keller Wirtschaft.* 12:347-76
  77. Gartel, W. 1972. Die Reblattgallmilbe *Eriophyes vitis* PGST. der Erreger der Pockenkrankheit (Erinose) als Knospenschädling und als Ursache starken Blattrollens. *Weinbau Kellerwirtschaft.* 19:589-614
  78. Guennelon, G., d'Arcier, F. 1972. Pigeage sexuel de l'eudemis de la vigne (*Lobesia botrana* Schiff.) dans la région d'Avignon. *Rev. Zool. Agric. Pathol. Veg.*
  79. Giannotti, J., Caudwell, A., Vago, C., Duthoit, J. L. 1969. Isolement et purification de micro-organismes de type mycoplasme à partir de vignes atteintes de flavescence dorée. *C.R. Acad. Sci. Paris, Sér. D* 268:845-47
  80. Günyadin, T. 1974. A survey of vine pests in south-east and east Anatolia. *Diyarb. Bölge Ziraat Muc. Inst.* 42:170
  81. Hatzinikolis, E. N. 1970. Neuf espèces d'acariens signalées pour la première fois en Grèce. *Ann. Inst. Phytopath. Benaki N.S.* 9(3):238-41
  82. Hewitt, W. et al. 1958. Transmission of fanleaf virus by *Xiphinema index*. *Phytopathology* 48:293
  83. Hewitt, W. B., Raski, D. J. 1967. Facteurs limitant la production 6:La vigne *Span* 10(1):56-59
  84. Hoffman, A. 1962. See Ref. 19, Vol 1, 2:902-3
  - 84a. Huffaker, C. B. 1971. *Biological Control*, pp. 273-274. New York: Plenum. 511 pp.
  85. Hurpin, B. 1962. See Ref. 19, Vol. 1, 1:24-122
  86. Ignator, B., Kirkov, K. 1972. The grape vine weevils. *Rastch. Zash.* 20(1):29-31
  87. Iordanou, N. 1974. Chemical control of grape berry moth *Tech. Pap. Agric. Res. Inst. Cyprus* 5:7
  88. Iren, Z. 1972. Investigations to determine the most important pests of viticulture in central Anatolia. *Anka Bölge Zorai Mucad. Enstit.* 40-41:168-69
  89. Ishii, K. 1975. Control of grape disease and insect pests in Japan. *Jpn. Pestic. Inf.* 23:17-23
  90. Ivan, A., Zahatia, V. 1969. New aspects of attack produced by *Agriotles lineatus* on vines *Rev. Hort. Vitic.* 6:66-69
  91. Ivanov, I. 1969. An experiment in forecasting and the control of the one-banded grape moth in the Burgas district. *Rastch. Zash.* 17(8/9):30-38
  92. Ivanov, I. 1969. On the biology of the one-banded grape moth. *Rastch. Zash.* 17(12):19-23
  93. Jensen, F. L. 1969. Microbiol insecti-

- cides for control of grape leaf folder. *Calif Agric.* 23(4):5-6
94. Jensen, F. L., Flaherty, D. D., Chiarrappa, L. 1969. Population densities and economic injury levels of grape leafhoppers. *Calif. Agric.* 23(4):9-10
  95. Jubb, G. L., Cox, J. A. 1974. Catches of redbanded leafroller moths in liquid bait traps in Erie County, Pennsylvania vineyards:12 years summary. *J. Econ. Entomol.* 67(3):448-49
  96. Jubb, G. L. 1973. Catches of *Episimus argutanus* (Clem) in grape berry moth sexpheromone traps in Pennsylvania. *J. Econ. Entomol.* 66(6):1345-46
  97. Kara'ozova, A. 1971. The control of the grape moth in 1970. *Rastch. Zash.* 19(5):13-16
  98. Kazas, I. A., Gorkavenko, A. S., Kiryukhin, G. A., Asriev, E. A. 1971. The protection of vineyards from *Phylloxera*. *Vsesay Nauch. Protcolvof. Stant. Odessa.* 264 pp.
  99. Kazas, I., Gorkavenko, A., Kiryukhin, G. A., Poldenko, V. 1966. *Le Phylloxera de la Vigne Simferopol (Crimée).* 157 pp.
  100. Khalilov, B. B. 1971. Vine entomofauna in Azerbaijan. *C.R. Int. Cong. Entomol. Moscow, 13th, 1968,* 2:344-45
  101. Kharizanov, A. 1969. Conclusions from the control of the variegated vine moth in the Plovdiv district in 1968. *Rastch. Zash.* 17(6):20-22
  102. Kharizanov, A. 1969. A new pest of grape vine in Bulgaria. *Rastch. Zash.* 17(11):21-23
  103. Kharizanov, A., Stoilov, A. 1969. The grape-vine mite and its control in the Plovdiv district. *Rastch. Zash.* 17(4): 25-28
  104. Kinn, D. N., Doult, R. L. 1972. Initial survey of arthropods found in North Coast vineyards of California. *Environ. Entomol.* 1(4):508-13
  105. Kinn, D. N., Doult, R. L. 1972. Natural control of spider mites on wine grape varieties in Northern California. *Environ. Entomol.* 1(4):513-18
  106. Kisakurek, O. R. 1972. Studies on the distribution, rate of infestation, parasites and predators of the grape cluster moth (*Lobesia botrana* Den. and Schiff.) in southern districts of Anatolia. *Bitki Kor. Bült.* 12(3):183-86
  107. Kiskin, P. K. 1961. Regeneration des racines et résistance au Phylloxera. *Acaad. Sci. Moldavie Kichinev. Inst. Zool. Publ.* 1:71-81
  108. Klerk, C. A. de 1974. Biology of *Phylloxera vitifoliae* (Fitch) in South Africa. *Phytophylactica* 6:109-18
  109. Kulkarni, K. A. 1971. Bionomics of the grape flea beetle *Sceledonta strigicollis* (Motsch.). *Mysore J. Agric. Sci.* 5: 308-16
  110. Lafon, J., Couillaud, P., Hude, R. 1955. *Maladies et parasites de la vigne.* Paris: Bailliere. 364 pp.
  111. Laing, J. E., Calvert, D. L., Huffaker, C. B. 1972. Preliminary studies of effects of *Tetranychus pacificus* McG. on yield and quality of grapes in the san Joaquin Valley California. *Environ. Entomol.* 1(5):658-63
  112. Laurent, M. 1972. Lutte contre les Tordeuses de la Grappe. *Vignes Vins Janv. Fév.* 206:20-30
  113. Liebermann, J., Espul, J. C., Mansur, P. S. 1971. On *Miogryllus convolutus* Johannson in the vineyards of San Carlos, La Consulta, Mendoza. *Idia* 281:55-61
  114. Lipetskaia, A. D., Rouzaiev, K. C. 1958. Les ravageurs et les maladies de la vigne. Moscow: 280 pp.
  115. Lynn, C. D. 1969. Omnivorous leafroller an important new grape pest in San Joaquin Valley. *Calif. Agric.* 23(4): 16-17
  116. Mac Lellan, C. R. 1973. Natural enemies of the light brown apple moth *Epiplatyas postvittana* (Wlk.) in the Australian capital territory. *Can. Entomol.* 105(5):681-700
  117. Maillat, P. 1957. Contribution à l'étude de la biologie du Phylloxera de la vigne. *Ann. Sci. Natl. Zool.*: 283-410
  118. Maillat, P. 1957. Le Phylloxéra de la vigne. Quelques faits biologiques et les problèmes qu'ils soulèvent. *Rev. Zool. Agric. Appl.* 7-9:1-19
  119. Mal'chenkova, N.I. 1970. *Eriophyes vitigineusgemma* sp.n., a pest of grape vine. *Zool. Zh.* 49(11):1728-31
  120. Mal'chenkova, N. I. 1971. On the mite fauna of the grape vine. See Ref. 201, pp. 107-21
  121. Manolache, C., Pasol, P., Romascu, E., Iordan, P., Naum, A., Sadagorschi, D., Popescu, M. 1974. Ecological contributions to the study of grape vines on sandy soils of the Platonesti-Saveni-Suditi area (*Ialomita*). *Ann. Inst. Cerc. Pentru Prot. Plant.* 10:257-64
  122. Mayet, V. 1890. *Les insectes de la vigne.* Paris:Masson. 466 pp.
  123. McGrew, J. R., Still, G. W. 1972. Control of grape diseases and insects in the eastern United States. *Farm. Bull.* 1893. 24

124. McKenzie, L. M., Beirne, B. P. 1972. A grape leafhopper *Erythroneura ziczac* Walsh and its Mymarid egg parasite in Okanagan Valley, British Columbia. *Can. Entomol.* 104(8):1229-33
125. McLachlan, R. A. 1970. Grape pest control in Queensland. *Queensl. Agric. J.* 96(4):231-35
126. Mikhailuk, I. B. 1970. The pests of grape vine. *Rastch. Zash.* 15(6):34-35
127. Mirica, A., Savescu, A., Mirica, I. 1969. Factors in forecasting for the control of the vine mealybug (*Pulvinaria betulae* L.). *Ann. Inst. Cerc. Pentru Protect. Plant.* 5:387-93
128. Moutous, G., Fos, A. 1971. Essais de lutte chimique contre la cicadelle de la vigne *Empoasca flavescens* Fabr. Résultats 1970. *Rev. Zool. Agric. Pathol. Vég.* 70(2):48-56
129. Moutous, G., Fos, A. 1971. Observations sur quelques ravageurs nouveaux ou occasionnels de la vigne. *Phytoma* 23(233):25-26
130. Moutous, G., Fos, A. 1972. La lutte contre la flavescence dorée et les cicadelles de la vigne. *Rev. Zool. Agric. Pathol. Vég.* 71(1):55-60
131. Moutous, G., Fos, A. 1973. Influence of populations levels of the vine Cicadellid (*Empoasca flavescens* Fab.) on the "scorching" symptoms on the leaves. *Ann. Zool. Ecol. Anim.* 5(2):173-85
132. Myburgh, A. C., Whitehead, V. B., Dairber, C. C. 1973. Pests of deciduous fruit, grapes and miscellaneous other horticultural crops in South Africa. *Entomol. Mem. Dep. Agric. Tech. Serv. Rep. S. Africa* 27. 38 pp.
133. Neamtu, I., Varna, P., Bahnareanus, M. 1969. Considerations regarding the attack of the vine moth on vines in vineyards at Husi. *Rev. Horticult. Vitic.* 6:63-65
134. Nedov, P. N. 1971. A study of the resistance of the grape vine to *Phylloxera vastatrix* Planch. *C.R. Int. Congr. Entomol. Moscow, 13th, 2-9 Août 1968* 2:366
135. Nikolaev, P. I., Ass, M. Ya. 1973. Details of the development of the vine phylloxera (*Dactylosphaera vitifolii* Fitch) on the Sochi coast of the Black Sea. *Byull. Mosk. Obs. Ispyt. Prir. Otdel Biol.* 78(2):5-20
136. Odak, S. C., Dhamdhare, S. V. 1970. New record of Hymenopterous parasites of *Sylepta lunalis* Guen, a pest of grapevines. *Indust. J. Entomol.* 32(4):395
137. Olalquiaga Faure, G., Contesse Pinto, J. 1959. Pests of grape vine in Chile. *FAO Plant. Protect. Bull.* 7(6):73-77
138. Panis, A. 1975. Lutte contre la cochenille farineuse dans le vignoble méditerranéen. *Prog. Agric. Vitic.* 92(15/16):470-73
139. Pantanelli, E. 1911. Danni di thrips sulle vite americane. *St. Sperim. Agric. ital.* 44(7):469-514
140. Pinnock, D. E., Mrlstead, J. E., Coe, N. F., Stegmiller, F. 1973. Evaluation of *Bacillus thuringiensis* formulations for control of larvae of western grape leaf skeletonizer. *J. Econ. Entomol.* 66(1):194-97
141. Plaut, H. N. 1973. On the biology of *Paropta paradoxus* (H.S.) on grape vine in Israel. *Bull. Entomol. Res.* 63(2):237-45
142. Predescu, S., Petranau, F., Lazar, S. 1969. Contributii la studiul componentelor biochimice a insectei *Sparaganothis pilleriana* Schiff. si a compozitei chimice a vitei de vi ca mediu de hrana. *Sucr. Stintif. Inst. Agric. Timis. Agron* 12:437-46
143. Prota, R. 1970. Un nuovo insetto dannoso ai frutti feri della Sardegna : la *Ceresa bubalus* F. *Studi Sassar.* 18(3):48-56
144. Purcell, A. H. 1975. Role of the blue-green sharpshooter, *Hordnia circellata*, in the epidemiology of Pierce's disease of grapevines. *Environ. Entomol.* 4(5):745-52
145. Pykhova, V. T. 1968. The control of the grape moth. *Rastch. Zash.* 13(11):48-49
146. Rambier, A. 1951. A propos du *Vesperus xartati* Muls. *Prog. Agric. Vitic.* No. 7-8, pp. 89-93
147. Rambier, A. 1969. Influence de traitements au DDT, au parathion et au carbaryl sur les ennemis naturels de *Panonychus ulmi* (Koch). *C. R. Symp. OILB Lutte Intég. Vignes, 4th*, pp. 173-78
148. Rambier, A. 1972. Les acariens dans le vignoble. *Prog. Agric. Vitic.* 89(16):385-97
149. Richard, M. 1975. Compte rendu des essais de lutte contre la pyrale. *Vigne Champ.* 1:18-20
150. Rings, R. W. 1972. Contributions to the bionomics of climbing cutworms distribution and developmental biology of *Agrotis barnesi*. *J. Econ. Entomol.* 65(2):397-401
151. Rings, R. W. 1972. Contributions to the bionomics of climbing cutworms; the

- distribution and developmental biology of the brown climbing cutworm, *Rhynchagrotis cupida*. *J. Econ. Entomol.* 65(3):734-37
152. Rivnay, E. 1939. Studies in the biology and ecology of *Retithrips syriacus* Mayet with special attention to its occurrence in Palestine. *Bull. Soc. Fouad Ier Ent. Cairo*, pp. 150-82
  153. Roehrich, R. 1970. Essais de deux produits commerciaux à base de *Bacillus thuringiensis* Berl. pour la protection de la vigne contre l'Eudemis *Lobesia botrana* Schiff. *Rev. Zool. Agric. Pathol. Veg.* 69(4):74-78
  154. Sampayo Fernandez, M., Hernandez Esteruelas, P. 1973. The vine moth (*Lobesia botrana* Schiff.). *Bol. Inf. Plagas* 108:7-11
  155. Sarai, D. S. 1972. Seasonal history and effect of soil moisture on mortality of newly hatched larvae of the grape root borer in Southern Missouri. *J. Econ. Entomol.* 65(1):182-84
  156. Saxena, D. K. 1970. Honey bees and wasps as pests of grape. *J. Bombay Nat. Hist. Soc.* 67(1):121-22
  157. Schruft, G. 1972. Les Tydeidé (Acariens) sur vigne. *Bull. OEPP* 3:51-55
  158. Schruft, G. 1972. Effects secondaires de fongicides agissant sur les Acariens sur vigne. *Bull. OEPP* 3:57-63
  159. Schruft, G. 1972. Das Vorkommen von Milben aus der Familie Tydeidae (Acari) an Reben. VI Beitrag über Untersuchungen zur Faunistik und Biologie der Milben (Acari) an Kulturreben (*Vitis* sp.). *Z. Angew. Entomol.* 71(2):124-33
  160. Schwester, D. 1972. Cicadelles de la vigne. *Bull. OEPP* 3:37-42
  161. Schwester, D., Carle, P., Moutous, G. 1961. Sur la transmission de la flavescence dorée des vignes par une cicadelle. *C. R. Seances Acad. Agric. Fr.* 147(18):1021-24
  162. Schwester, D., Moutous, G., Bonfils, J., Carle, P. 1962. Etude biologique des cicadelles de la vigne dans le sudouest de la France. *Ann. Epiph.* 13(3):205-37
  163. Sengalevich, G. 1969. New pests on permanent plantations in the Plovdiv area. *Rastch. Zash.* 17(3):21-23
  164. Servicio Para el Agricultor. 1973. La avispa de la uva. *Notas Agric.* 6(26):105
  165. Seryi, N. I. 1971. On the biology of the vine moth (*Clysia ambiguella* H. B.). See Ref. 201, pp. 86-90
  166. Seryi, N. I. 1972. The biological principles of methods for the control of the pests of grape vines in Moldavia. See Ref. 201, pp. 95-103
  167. Sidhu, A. S., Singh, G. 1972. Studies on the chemical control of *Oligonychus mangiferus* (Rahman and Sapra) on grape vine. *J. Res. Punjab Agric. Univ.* 8(4):462-65
  168. Silvestri, F. 1939. Compendio di entomologia applicata. *Ed. Portici* Vol. 1, 2. 974., 685 pp.
  169. Siriez, H. 1970. Un ravageur presque oublié : la pyrale de la vigne. *Phytoma* 22(215):41-47
  170. Smith, L. M., Stafford, E. 1955. Grape pests in California. *Calif. Agric. Exp. Stn. Circ.* 445. 63 pp.
  171. Sohi, A. S., Singh, S. 1970. New pests of grape vine. *Labdev. J. Sci. Technol.* 8 B 3:170
  172. Sorauer, P. 1949-1958. *Handbuch der Pflanzenkrankheiten.*, Vol. 1-6. Berlin: Parey
  173. Stafford, E. M., Kido, H. 1969. Newer insecticides for the control of grape insect and spider mite pests. *Calif. Agric.* 23(4):6-8
  174. Stellwaag, F. 1928. *Die Weinbauinsekten der Kulturländer.* Berlin : Parey 884 pp.
  175. Stevenson, A. B. 1962. Insecticide dips to control grape phylloxera on nursery stocks. *J. Econ. Entomol.* 55(5):804-5
  176. Stevenson, A. B. 1964. Seasonal history of root-infesting *Phylloxera vitifoliae* (Fitch) in Ontario. *Can. Entomol.* 96(7):979-87
  177. Stevenson, A. B. 1966. Seasonal development of foliage infestations of grape in Ontario by *Phylloxera vitifoliae* (Fitch). *Can. Entomol.* 98(12):1299-305
  178. Stevenson, A. B. 1968. Soil treatments with insecticides to control the rooform of the grape Phylloxera. *J. Econ. Entomol.* 61(5):1168-71
  179. Stevenson, A. B. 1970. Endosulfan and other insecticides for control of the leaf form of the grape phylloxera in Ontario. *J. Econ. Entomol.* 63(1):125-28
  180. Stevenson, A. B. 1970. Strains of the grape phylloxera in Ontario with different effects on the foliage of certain grape cultivars. *J. Econ. Entomol.* 63(1):135-8
  181. Still, G. W. 1962. Cultural control of the grape berry moth. *ARS Agric. Inf. Bull.* 256:1-8
  182. Still, G. W., Rings, R. W. 1973. Insect and mite pests of grapes in Ohio. *Res. Bull. Ohio Agric. Res. Dev. Cent.* 1060. 30 pp.

183. Suire, J. 1954. Contribution à l'étude morphologique des Microlépidoptères de la vigne. *Ann. Ecole Natl. Agric. Montpellier* 24 (3 and 4):1-7
184. Tairov, V. E. 1967. *Questions Relatives à Laviticulture et à la Vinification*. (In Russian) Kiev : 386 pp.
185. Taschenberg, E. F., Carde, R. T., Roelofs, W. L. 1974. Sex pheromone mass trapping and mating disruption for control of redbanded leafroller and grape berry moth in vineyards. *Environ. Entomol.* 3(2):239-42
186. Troitzky, N. N. 1929. Beiträge zur kenntnis der Reblaus und der Reblauswiderstomdsfähigkeit der Weinrebe : Die Reblausfrage in Mittel-Europa Leningrad : Bur. Appl. Entomol. 191 pp.
187. Vega, E. 1972. Control de la filoxera de la vid con hexachlorobutadien. *Idia* 290:15-18
188. Vidal, J. P. 1974. Le piegeage de l'Eudemis (*Polychrosis botrana*) substances attractives. *Bull. Tech. Pyr. Or.* 72: 67-71
189. Vidano, C. 1957-58. Le Cicaline italiane della vite. *Boll. Zool. Agric. Bachic. Milan* 2(1):61-115
190. Vidano, C. 1959. Sulla identificazione specifica di alcuni *Erythroneurini* europei. *Ann. Mus. Civ. Stor. Natur. Giacomo Doria* 71:328-48
191. Vidano, C. 1962. La *Empoasca libyca* Bergevin nuovo nemico della vite in Italia. *Cent. Entomol. Alp. forest. CNR* 55:327-45
192. Vidano, C. 1963. Deviazione trofica ampelofila della *Ceresa bubalus* Fabricius e rispondenza reattiva del vegetale. *Atti. Accad. Sci. Torino* 98:193-212
193. Vidano, C. 1963. Appunti comparativi sui danni da cicaline alla vite. *Inf. Fitopatol.* 13:173-77
194. Vidano, C. 1965. Responses of Vitis to insect vector feeding. *Proc. Conf. Virus Vector, Univ. Calif, Davis* pp. 73-80
195. Viswanath, B. N., Nalawadi, U. G., Kulkarni, K. A. 1970. The yellow banded wasp, *Vespa cincta* Fabr. as a pest of grapes. *Agric. Res. J. Kerala* 8(1):53
196. Voigt, E. 1970. Influence of meteorological factors on the population dynamics of *Eupoecilia ambiguella* Hb and *Lobesia botrana* Den. and Schiff. *Növény. Korsz.* 4:63-78
197. Voigt, E., Bodor, J., Javor, A. 1973. Morphological descriptions of larvae of moth injurious to grapevines (Hungarian). *Novenynezesitesi Novenyter mesztesi Kut. Intez. Sopronhorpacs Kozl.* 6:51-64
198. Vuittenez, A. 1961. Les nématodes vecteurs de virus et le problème de la dégénérescence infectieuse de la vigne. In *ACTA, Les Nématodes*, Paris, pp. 55-77
199. Whitehead, V. B., Rust, D. J. 1972. Control of the fruit piercing moth *Serodes partita* F. *Phytophylactica* 4(1): 9-12
200. Winkler, A. S. 1959. Pierce's disease investigations. *Hilgardia* 19:207-264
201. Yaroshenko, M. F. 1972. The insect fauna of Moldavia and its economic importance. In *Izdatel'stvo Shtiintsa*. 142 pp.
202. Yousef, A. T. A. 1970. Mites associated with vine trees in the UAR. *Z. Angew. Entomol.* 67(1):1-6
203. Zinca, N. 1964. Cercetari asupra morfologiei, biologiei, si combaterii trip-sului vitei de vie *Anaphthrips vitis* Pr. *Inst. Cent. Cerc. Agric. Ann. Protect. Plant.* 2:299-305