

Approved Survey Methodology for Negative Data

The survey methodology presented on the [Approved Methods for Pest Surveillance](#) webpages lists the most up-to-date, approved methods for survey and identification/diagnostics of target pests on the Priority Pest List. The information on the webpage supersedes any survey and identification/diagnostic information found in any other document (e.g., Commodity-based Survey References and Guidelines, CAPS Pest Datasheets, etc.). All other documents will eventually be revised to include the information contained on the Approved Methods for Pest Surveillance webpage; however, **these pages will always be the authoritative source for the most up-to-date approved methods for Priority Pests.**

Click [here](#) for the 2021 Approved Methods for Pest Surveillance (AMPS).

PPQ CAPS Pest Datasheets

PPQ Science & Technology (S&T) CAPS Support develops pest datasheets for the Priority Pest List. The pest datasheets are designed as stand-alone files, and are available on the individual pest pages of the Approved Methods for Pest Surveillance site.

Visual Surveys Conducted in Nurseries

It was decided at the 2013 National CAPS Committee Meeting that surveys conducted in nurseries must follow the approved methods for target pests. Negative data will be allowed for a visual survey only if visual is an approved method for the target species. If nursery owners do not want traps hung in nurseries, surveyors can place traps near the nurseries or in right-of-ways.

Insects – Survey Considerations

For 2011 and beyond, negative data may be reported for these target pests **only** when surveyed for by the Approved Methods for Pest Surveillance method. For species with attractants and/or lures available, the approved trap type and lure **must** be used in order to report negative data.

If “visual” is listed as the only approved survey method for a target pest, then traps and lures have not yet been proven effective for attracting that target. Negative data may only be reported for these species by conducting a visual survey and not from any trap and lure combination.

Trap recommendations

The S&T CAPS Support Team compares published survey methods and confers with subject matter experts to determine which method is most effective for early detection surveys for each CAPS target. If there is more than one effective trap, the team narrows the options to the best trap(s) in order to establish homogeneity in the dataset. Decisions are made based on the scientific data available, cost, and ease of use (for both surveyors and identifiers).

S&T CAPS Support welcomes your input. If you want CAPS to consider a different survey method, please send the evidence supporting the method (literature, personal communication,

etc.) to STCAPS@usda.gov with the subject line “suggested approved method for *Pest name*”. **This must be done in advance of using these survey methods in surveys. You may not enter data** from survey methods that have not yet been approved.

Ordering traps and lures

Unless otherwise noted, all traps and lures should be purchased through the IPHIS Survey Supply Ordering System. The trap and lure product names on the Approved Methods for Pest Surveillance pages are synchronized with the product names in the IPHIS Survey Supply Ordering System.

IMPORTANT: Unless noted otherwise, when more than one lure is listed, all of the lures are required to report negative data for that species. For example, for *Tetropium castaneum*, three lures are listed:

- Spruce Blend Lure
- Geranyl Acetol Lure
- Ethanol Lure

All three lures are required for negative data reporting for this target.

In some cases, there are multiple lure options that may be used. These will be designated by the **Option** header (e.g., Option 1, Option 2). It is rare that there is more than one lure option. When multiple lure options are listed, additional guidance is included to assist states in determining which lure to use.

In some cases, multiple traps or colors of traps are listed as approved. Additional guidance is included to assist states in determining which trap to use.

Example for *Adoxophyes orana*:

- Paper Delta Trap, 2 sticky sides, Brown
- Paper Delta Trap, 2 sticky sides, Green
- Paper Delta Trap, 2 sticky sides, Orange

Any one of these traps would be equally acceptable for use. For this insect, there is a **Method Note** re-affirming this, “Trap color is up to the State and does not affect trap efficacy.”

IMPORTANT: Do not place lures for two or more target species in a trap unless otherwise recommended. Effects of lures on non-target species cannot be predicted based solely on pheromone chemistry, taxonomic relationship, etc. Because of this, lures should not be combined in individual traps unless PPQ has specific data indicating that the trap will remain a functional detection tool for both species.

Some trap and lure combinations are approved for more than one target. For exotic wood borers and bark beetles, see the table in [EWB/BB Survey Tables](#) on the [Exotic Wood Borer / Bark Beetle Survey Reference](#) page.

Trap spacing

In general, when trapping for more than one species of moth, the traps for different moth species should be spaced at least 20 meters (65 feet) apart. When trapping for wood borers or bark beetles, separate traps with different lure combinations by at least 30 meters (98 feet).

Insects – Identification Information

For the majority of insects, morphology is the basis for identification. In a few instances, molecular techniques are used for confirmation. Please refer to the individual pest pages and datasheets for specific information.

The “Easily Confused Species” section lists families, genera, and species that may be confused with the target species. This information is not to be interpreted as an exhaustive list of all species that could be confused for the target.

Mollusks – Survey Considerations

All mollusk surveys will be by visual inspection. Use the cited references for additional information on identifying high risk sites, seasonality and time of day to survey, and signs of mollusk presence.

[Data Entry Guide for Selected Taxonomic Groups](#) should be used to determine the appropriate requirements for reporting negative data.

Mollusks – Identification Information

See specific information on the pest datasheets.

Nematodes – Survey Considerations

Negative data may be reported for these target pests only when surveyed for by the Approved Methods for Pest Surveillance. For most nematodes, surveys will be conducted via soil sampling. If plant hosts are present, sampling host roots or visual sampling can be used in conjunction with soil sampling. When more than one survey method is listed, all methods can be used depending on the type of survey. The methods, however, are listed in order from most preferred to least preferred. Please refer to the individual pest pages and datasheets for detailed survey information.

[Data Entry Guide for Selected Taxonomic Groups](#) should be used to determine the appropriate requirements for reporting negative data.

Nematodes – Identification Information

Negative data may be reported for these target pests only when using the identification method listed in the Approved Methods for Pest Surveillance. These methods correspond to those commonly used to confirm a nematode identity. Any molecular methods that have been validated by the S&T lab in Beltsville, MD also are included in the Approved Methods for Pest Surveillance. Work instructions are available for each method validated by S&T-Beltsville. If more than one diagnostic method is listed, a combination of methods may be required to confirm a nematode identity.

For the majority of nematodes, morphology is the basis for identification. In a few instances, biochemical or molecular techniques are used for confirmation. Please refer to the individual pest pages and datasheets for specific information.

The "Easily Confused Species" section lists nematodes and conditions that may be confused with the target species. This information is not to be interpreted as an exhaustive list of all species that could be confused for the target.

Information is provided on diagnostic methods from the scientific literature in individual pest datasheet and in the Approved Methods for Pest Surveillance pest information page in the "Literature-based Methods/Diagnostics" section. Although these methods have not been validated by PPQ, many labs and state institutions may find the information useful when making preliminary pest identifications.

Pathogens – Survey Considerations

Negative data may be reported for these target pests only when using the identification or diagnostic method listed in the Approved Methods for Pest Surveillance. For most pathogens, surveys will be conducted visually by looking for characteristic symptoms (an indication of disease by reaction of the host, e.g. canker, leaf spot, wilt, etc.) and/or signs (indication of disease from direct observation of a pathogen or its parts). When more than one survey method is listed, all methods can be used depending on the type of survey. The methods, however, are listed in order from most preferred to least preferred. Please refer to the individual pest pages and datasheets for detailed survey information.

Pathogens – Diagnostic Information

Negative data may be reported for these target pests only when using the identification or diagnostic method listed in the Approved Methods for Pest Surveillance. These methods correspond to those commonly used to confirm a pathogen identity. Any molecular methods that have been validated by the S&T lab in Beltsville, MD also are included in the Approved Methods for Pest Surveillance. Work instructions are available for each method validated by

S&T-Beltsville. If more than one diagnostic method is listed, a combination of methods may be required to confirm a pathogen's identity.

For the many pathogens, morphology is the basis for identification. Serology or molecular techniques are often used for confirmation as well. Please refer to the individual pest pages and datasheets for specific information.

The "Easily Confused Species" section lists pathogens and conditions that may be confused with the target species. This information is not to be interpreted as an exhaustive list of all species that could be confused for the target.

Information is provided on diagnostic methods from the scientific literature in individual pest datasheet and in the Approved Methods for Pest Surveillance pest information page in the "Literature-based Methods/Diagnostics" section. Although these methods have not been validated by PPQ, many labs and state institutions may find the information useful when making preliminary pest identifications.

Weeds – Survey Considerations

All weed surveys will be by visual inspection. Please see detailed survey information by clicking on the target species' datasheet on the main table of the Approved Methods for Pest Surveillance.

Weeds – Identification Information

At this time, all weed identification will involve a morphological confirmation by a botanist. See specific information on the pest pages and datasheets.

For further information contact:

Heather Moylett

919-855-7428

heather.moylett@usda.gov

S&Tcaps@aphis.usda.gov